

TITLE:


METHODS AND COMPOSITIONS FOR
SORTING AND/OR DETERMINING ORGANISMS

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CERTIFICATION UNDER 37 C.F.R. § 1.10

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Title Of The Invention:

Methods And Compositions For Sorting And/Or Determining Organisms

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Cross Reference to Related Applications:

This application claims the benefit of U.S. Provisional Application No. 60/250,930 filed on November 30, 2000.

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Background of the Invention:1. Field of the Invention

This invention is related to the field of combined probe-based and binding partner-based detection, analysis and/or quantitation of organisms and/or cells.

2. Description of the Related Art

Nucleic acid hybridization is a fundamental process in molecular biology. Probe-based assays are useful in the detection, quantitation and/or analysis of nucleic acids. Nucleic acid probes have long been used to analyze samples for the presence of nucleic acid from bacteria, fungi, virus or other organisms and are also useful in examining genetically-based disease states or clinical conditions of interest. Nonetheless, nucleic acid probe-based assays have been slow to achieve commercial success. This lack of commercial success is, at least partially, the result of difficulties associated with specificity, sensitivity and/or reliability.

Despite its name, Peptide Nucleic Acid (PNA) is neither a peptide, a nucleic acid nor is it an acid. Peptide Nucleic Acid (PNA) is a non-naturally occurring polyamide that can hybridize to nucleic acid (DNA and RNA) with sequence specificity (See: United States Patent No. 5,539,082 and Egholm et al., *Nature* 365: 566-568 (1993)). In point of fact, PNA has been consistently characterized as a nucleic acid mimic rather than a nucleic acid analog since it is not derived from nucleic acid or its component nucleotides or nucleosides as well as comprises unique properties (See: Nielsen, P.E., *Acc. Chem. Res.* 32: 624-630 (1999)). Being a non-naturally occurring molecule, unmodified PNA is not known to be a substrate for the enzymes that are known to degrade peptides or nucleic

acids. Therefore, PNA should be stable in biological samples, as well as have a long shelf-life. Unlike nucleic acid hybridization, which is very dependent on ionic strength, the hybridization of a PNA with a nucleic acid is fairly independent of ionic strength and is favored at low ionic strength, conditions that strongly disfavor the hybridization of nucleic acid to nucleic acid (Egholm et al., *Nature*, at p. 567). Because of their unique properties, it is clear that PNA is not the equivalent of a nucleic acid in either structure or function. Consequently, PNA probes need to be evaluated for performance and optimization to thereby confirm whether they can be used to specifically and reliably detect a particular nucleic acid target sequence, particularly when the target sequence exists in a complex sample such as a cell, tissue or organism.

Like nucleic acid probes, antibodies have been used to examine samples for the purpose of determining organisms. Labeled nucleic acid probes used in combination with labeled antibodies have also been used to identify bacteria by flow cytometric analysis. (See: Wallner et al., *System. Appl. Microbiol.*, 19: 569-576 (1996)). Importantly, Wallner et al. analyzed the bacteria as stained by the labeled nucleic acid probes and labeled antibodies but did not immobilize the bacteria to a particle, bead or other solid carrier for analysis.

Luminex (Austin, Texas) has recently introduced coded beaded supports and an instrument suitable for determining the coded beaded supports as well as one or more reporter moieties bound thereto. (See: Luminex Product Literature) The coded beaded supports incorporate a proprietary, precision process to internally dye same-sized polystyrene microspheres with two fluorophores. *Id.* Using precise ratios of the two fluorophores, Luminex has created 100 different coded microsphere sets wherein each set is distinguished based on its internal dye ratio using the Luminex 100™. *Id.* In addition to determining the code of each bead, the Luminex 100™ can also determine a reporter moiety on the beads with highly accurate quantitation. *Id.*

By providing coded beaded supports having surface carboxyl groups or Lumavidin™, Luminex proposes that their coded beaded supports and instruments can be adapted for applications involving Molecular Biology, Immunoassays, Enzymatic Assays & Reporter-Ligand Assays, depending on the nature of the immobilized ligands. *Id.* Importantly, Luminex does not teach or suggest the integration of specific analyte capture with specific analyte staining as a means to achieve improved discrimination in

the assay or certainty of the result and in particular, Luminex does not appear to teach or suggest that organisms of any kind can be bound to and determined using their beads and/or instrument. In fact, Applicants are not aware of any example of using a specific binding partner to immobilize, for determination, specifically stained organisms of

5 interest to a solid carrier.

Summary of the Invention:

This invention is generally directed to methods and compositions that pertain to the use of molecular probes, for the selective staining of organisms or cells, in combination with the selective capture of organisms or cells using binding partners. The methods and compositions of this invention can be used for sorting and/or determining an organism or organisms of interest. When sorting is chosen, sorting can be performed, for example, either by the use of coded beaded supports or by the use of an array. Because selectivity can be affected at two very different levels of molecular recognition, the methods and compositions of this invention provide for enhanced assay discrimination and/or enhanced certainty of the result.

The utility of the methods and compositions of this invention can be further enhanced by the use of independently detectable molecular probes that facilitate the multiplexing of the staining process. Multiplexing of the methods and compositions is also facilitated at the immobilization step of the process by, for example, the use of different binding partners that are used for the selective immobilization of different organisms of interest. Multiplexing an assay at the immobilization step can also be facilitated by the use of coded beaded supports wherein the "code" for the different beads is associated with the sample source or with another parameter of interest. Hence, the methods and compositions of this invention facilitate a broad scope of flexibility of analysis in a way that overcomes numerous limitations of the prior art

Brief Description of the Drawings

Figure 1A is a print of a composite digital image of the red and green images taken with a microscope equipped with a CCD camera. The red spheres in the image are coded beaded supports and the green rods are *Salmonella choleraesuis* bacteria immobilized to the surface of the beads.

Figure 1B is a print of a composite digital image of the red and green images taken with a microscope equipped with a CCD camera. The red sphere in the image is a coded beaded support and the green rods are *Listeria monocytogenes* bacteria that do not appear to bind to the surface of the bead.

Figure 2A is a print of a composite digital image of the red and green images taken with a microscope equipped with a CCD camera. The red sphere in the image is a coded beaded support and the green rods are *Salmonella choleraesuis* bacteria immobilized to the surface of the beads.

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Figure 2B is a print of a composite digital image of the red and green images taken with a microscope equipped with a CCD camera. The red spheres in the image are coded beaded supports and the green rods are *Listeria monocytogenes* bacteria immobilized to the surface of the beads.

Detailed Description of The Invention

1. Definitions:

For the purposes of interpreting of this specification the following definitions shall apply and whenever appropriate, terms used in the singular shall also include the plural and vice versa.

a. As used herein, a “nucleobase” means those naturally occurring and those non-naturally occurring heterocyclic moieties commonly known to those who utilize nucleic acid technology or utilize peptide nucleic acid technology to thereby generate polymers that can sequence specifically bind to nucleic acids.

b. As used herein, a “nucleobase sequence” means any segment of a polymer that comprises nucleobase-containing subunits. Non-limiting examples of suitable polymers or polymers segments include oligodeoxynucleotides (e.g. DNA), oligoribonucleotides (e.g. RNA), peptide nucleic acids (PNA), nucleic acid analogs, nucleic acid mimics, and/or chimeras.

c. As used herein, a “target sequence” is the nucleobase sequence of a nucleic acid that is found in an organism of interest and to which a molecular probe is designed to hybridize sequence specifically thereto.

d. As used herein, a “nucleic acid” is a nucleobase sequence-containing polymer, or polymer segment, having a backbone formed from nucleotides, or analogs thereof.

e. As used herein, a “non-nucleic acid” is a nucleobase sequence containing polymer, or polymer segment, having a backbone formed from subunits that are not nucleotides, or analogs thereof. Peptide nucleic acids are a preferred non-nucleic acid polymer.

f. As used herein, the term “probe” or “molecular probe” means a nucleic acid or non-nucleic acid polymer (e.g. a DNA, RNA, PNA, nucleic acid analogs, nucleic acid mimics, chimera or linked polymer) having a probing nucleobase sequence that is designed to sequence specifically hybridize to a target sequence of a target molecule of an organism of interest.

g. As used herein, a “detectable molecular probe” is a probe or molecular probe that is detectable by instrument or method. For the avoidance of doubt, a “detectable molecular probe” need not be directly labeled with a detectable moiety (See: the

subsection entitled: "Unlabeled Molecular Probes", below for a discussion of determining unlabeled molecular probes).

h. As used herein, the term "antibody" means an antibody or antibody fragment that is capable of participating in an antibody/antigen binding interaction.

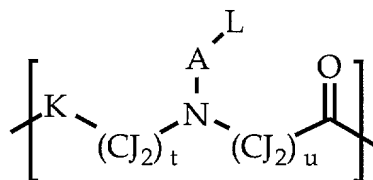
5 i. As used herein, the term "detectable antibody" means an antibody or antibody fragment that is detectable by instrument or method. For the avoidance of doubt, a detectable antibody need not be directly labeled with a detectable moiety since, for example, the antibody may be detected using a secondary antibody that is labeled with a detectable moiety.

10 j. As used herein, "stained" means that individual organisms are directly or indirectly marked with a detectable moiety as a result of the sequence specific hybridization of one or more detectable molecular probes to a target sequence within the organism.

15 k. As used herein, the term "peptide nucleic acid" or "PNA" means any oligomer, polymer, linked polymer or chimeric oligomer, comprising two or more PNA subunits (residues), including any of the polymers referred to or claimed as peptide nucleic acids in United States Patent Nos. 5,539,082, 5,527,675, 5,623,049, 5,714,331, 5,718,262, 5,736,336, 5,773,571, 5,766,855, 5,786,461, 5,837,459, 5,891,625, 5,972,610, 5,986,053 and 6,107,470; all of which are herein incorporated by reference. The term "peptide nucleic acid" or
20 "PNA" shall also apply to polymers comprising two or more subunits of those nucleic acid mimics described in the following publications: Lagriffoul et al., *Bioorganic & Medicinal Chemistry Letters*, 4: 1081-1082 (1994); Petersen et al., *Bioorganic & Medicinal Chemistry Letters*, 6: 793-796 (1996); Diderichsen et al., *Tett. Lett.* 37: 475-478 (1996); Fujii et al., *Bioorg. Med. Chem. Lett.* 7: 637-627 (1997); Jordan et al., *Bioorg. Med. Chem. Lett.* 7: 687-
25 690 (1997); Krotz et al., *Tett. Lett.* 36: 6941-6944 (1995); Lagriffoul et al., *Bioorg. Med. Chem. Lett.* 4: 1081-1082 (1994); Diederichsen, U., *Bioorganic & Medicinal Chemistry Letters*, 7: 1743-1746 (1997); Lowe et al., *J. Chem. Soc. Perkin Trans. 1*, (1997) 1: 539-546; Lowe et al., *J. Chem. Soc. Perkin Trans. 11*: 547-554 (1997); Lowe et al., *J. Chem. Soc. Perkin Trans. 1* 1:5 55-560 (1997); Howarth et al., *J. Org. Chem.* 62: 5441-5450 (1997); Altmann, K-H et al.,
30 *Bioorganic & Medicinal Chemistry Letters*, 7: 1119-1122 (1997); Diederichsen, U., *Bioorganic & Med. Chem. Lett.*, 8: 165-168 (1998); Diederichsen et al., *Angew. Chem. Int. Ed.*, 37: 302-305 (1998); Cantin et al., *Tett. Lett.*, 38: 4211-4214 (1997); Ciapetti et al., *Tetrahedron*, 53: 1167-

1176 (1997); Lagriffoule et al., *Chem. Eur. J.*, 3: 912-919 (1997); and the Peptide-Based Nucleic Acid Mimics (PENAMs) of Shah et al. as disclosed in WO96/04000.

In preferred embodiments, a PNA is a polymer comprising two or more subunits of the formula:



wherein, each J is the same or different and is selected from the group consisting of H, R¹, OR¹, SR¹, NHR¹, NR¹, F, Cl, Br and I. Each K is the same or different and is selected from the group consisting of O, S, NH and NR¹. Each R¹ is the same or different and is an alkyl group having one to five carbon atoms that may optionally contain a heteroatom or a substituted or unsubstituted aryl group. Each A is selected from the group consisting of a single bond, a group of the formula; -(CJ₂)_s- and a group of the formula; -(CJ₂)_sC(O)-, wherein, J is defined above and each s is a whole number from one to five. Each t is 1 or 2 and each u is 1 or 2. Each L is the same or different and is independently selected from the group consisting of J, adenine, cytosine, guanine, thymine, uridine, 5-methylcytosine, 2-aminopurine, 2-amino-6-chloropurine, 2,6-diaminopurine, hypoxanthine, pseudoisocytosine, 2-thiouracil, 2-thiothymidine, other naturally occurring nucleobase analogs, other non-naturally occurring nucleobases, substituted and unsubstituted aromatic moieties, biotin, fluorescein and dabcyI. In the most preferred embodiment, a PNA subunit consists of a naturally occurring or non-naturally occurring nucleobase attached to the aza nitrogen of the N-[2-(aminoethyl)]glycine backbone through a methylene carbonyl linkage.

l. As used herein, the terms "label" and "detectable moiety" are interchangeable and refer to moieties that can be attached to a molecular probe, antibody or antibody fragment to thereby render the molecular probe, antibody or antibody fragment detectable by an instrument or method.

m. As used herein, the term "chimera" or "chimeric oligomer" means a polymer comprising two or more linked subunits that are selected from different classes of subunits. For example, a PNA/DNA chimera would comprise at least two PNA subunits linked to at least one 2'-deoxyribonucleic acid subunit (For exemplary methods and

compositions related to PNA/DNA chimera preparation See: WO96/40709). Exemplary component subunits of the chimera are selected from the group consisting of PNA subunits, naturally and non-naturally occurring amino acid subunits, DNA subunits, RNA subunits and subunits of analogues or mimics of nucleic acids.

5 n. As used herein, the term “linked polymer” means a polymer comprising two or more polymer segments that are linked by a linker. The polymer segments that are linked to form the linked polymer are selected from the group consisting of an oligodeoxynucleotide (DNA), an oligoribonucleotide (RNA), a peptide, a polyamide, a peptide nucleic acid (PNA) and a chimera.

10 o. As used herein, the term “binding partner” means those molecules that bind to one or more other molecules in a specific manner. Because the binding partner interactions are specific, there is a degree of selectivity that is achieved depending on the nature of the binding partners chosen. Non-limiting examples of binding partner complexes (formed from the component binding partners) include antibody/antigen interactions, nucleic acid/nucleic acid interactions, enzyme/substrate interactions and receptor/ligand interactions. A non-limiting list of ligands includes avidin (and its analogs such as Streptavidin and LumavidinTM), lectins, carbohydrates, peptides and proteins. The preferred pair of binding partners used in the practice of this invention is the antibody/antigen.

15 p. As used herein, the term “solid carrier” means an object that has a surface that is broad enough to accommodate organisms linked thereto. Preferred materials used to construct the solid carrier include, but are not limited to, glass, quartz, plastic (e.g. polystyrene, polyamide, polyacrylic, polyethylene, polypropylene and PTFE (Teflon)) and gold. The preferred solid carriers are particles, beads, microscope slides, micro titre plates, membranes and arrays.

25 q. As used herein, an “array” is a two or three-dimensional object having one or more surfaces upon which two or more unique, identifiable locations are created. A microscope slide is one example of an object that can be used to manufacture an array.

30 2. Description Of The Invention

I. *General:*

Nucleic Acid Synthesis and Modification

Nucleic acid oligomer (oligonucleotide and oligoribonucleotide) synthesis has become routine. For a detailed description of nucleic acid synthesis please see *Gait, M. J., Oligonucleotide Synthesis: a Practical Approach. IRL Press, Oxford England.* Those of ordinary skill in the art will recognize that both labeled or unlabeled oligonucleotides (DNA, RNA and synthetic analogues thereof) are readily available. They can be synthesized using commercially available instrumentation and reagents or they can be purchased from commercial vendors of custom manufactured oligonucleotides. Patents that discuss various compositions, supports and methodologies for the synthesis and labeling of nucleic acids include: 5,476,925, 5,453,496, 5,446,137, 5,419,966, 5,391,723, 5,391,667, 5,380,833, 5,348,868, 5,281,701, 5,278,302, 5,262,530, 5,243,038, 5,218,103, 5,204,456, 5,204,455, 5,198, 540, 5,175,209, 5,164,491, 5,112,962, 5,071,974, 5,047,524, 4,980,460, 4,923,901, 4,786,724, 4,725,677, 4,659,774, 4,500,707, 4,458,066, and 4,415,732; all of which are herein incorporated by reference.

PNA Synthesis:

Methods for the chemical assembly of PNAs are well known (See: Patent Nos. 5,539,082, 5,527,675, 5,623,049, 5,714,331, 5,718,262, 5,736,336, 5,773,571, 5,766,855, 5,786,461, 5,837,459, 5,891,625, 5,972,610, 5,986,053 and 6,107,470; all of which are herein incorporated by reference (Also see: PerSeptive Biosystems Product Literature)). Chemicals and instrumentation for the support bound automated chemical assembly of peptide nucleic acids are now commercially available. Both labeled and unlabeled PNA oligomers are likewise available from commercial vendors of custom PNA oligomers. Chemical assembly of a PNA is analogous to solid phase peptide synthesis, wherein at each cycle of assembly the oligomer possesses a reactive alkyl amino terminus that is condensed with the next synthon to be added to the growing polymer. Because standard peptide chemistry is utilized, natural and non-natural amino acids are routinely incorporated into a PNA oligomer. Because a PNA is a polyamide, it has a C-terminus (carboxyl terminus) and an N-terminus (amino terminus). For the purposes of the design of a hybridization probe suitable for antiparallel binding to the target sequence (the preferred orientation), the N-terminus of the probing nucleobase sequence of the PNA probe is the equivalent of the 5'-hydroxyl terminus of an equivalent DNA or RNA oligonucleotide.

PNA Labeling:

Preferred non-limiting methods for labeling PNAs are described in US 6,110,676, WO99/22018, WO99/21881, WO99/49293 and WO99/37670, the examples section of this specification or are otherwise well known in the art of PNA synthesis and peptide synthesis.

5 Labels:

Non-limiting examples of detectable moieties (labels) suitable for directly labeling molecular probes, antibodies or antibody fragments used in the practice of this invention include a dextran conjugate, a branched nucleic acid detection system, a chromophore, a fluorophore, a spin label, a radioisotope, an enzyme, a hapten, an acridinium ester and a
10 chemiluminescent compound. Other suitable labeling reagents and preferred methods of attachment would be recognized by those of ordinary skill in the art of PNA, peptide or nucleic acid synthesis.

Preferred haptens include 5(6)-carboxyfluorescein, 2,4-dinitrophenyl, digoxigenin, and biotin.

Preferred fluorochromes (fluorophores) include 5(6)-carboxyfluorescein (Flu), 6-
15 ((7-amino-4-methylcoumarin-3-acetyl)amino)hexanoic acid (Cou), 5(and 6)-carboxy-X-rhodamine (Rox), Cyanine 2 (Cy2) Dye, Cyanine 3 (Cy3) Dye, Cyanine 3.5 (Cy3.5) Dye, Cyanine 5 (Cy5) Dye, Cyanine 5.5 (Cy5.5) Dye Cyanine 7 (Cy7) Dye, Cyanine 9 (Cy9) Dye (Cyanine dyes 2, 3, 3.5, 5 and 5.5 are available as NHS esters from Amersham,
20 Arlington Heights, IL), JOE, Tamara or the Alexa dye series (Molecular Probes, Eugene, OR).

Preferred enzymes include polymerases (e.g. Taq polymerase, Klenow PNA polymerase, T7 DNA polymerase, Sequenase, DNA polymerase 1 and phi29 polymerase), alkaline phosphatase (AP), horseradish peroxidase (HRP) and most
25 preferably, soy bean peroxidase (SBP).

Detectable and Independently Detectable Moieties/Multiplex Analysis:

In preferred embodiments of this invention, a multiplex hybridization assay is performed. In a multiplex assay, numerous conditions of interest are simultaneously or sequentially examined. Multiplex analysis relies on the ability to sort sample components
30 or the data associated therewith, during or after the assay is completed. In preferred embodiments of the invention, one or more distinct independently detectable moieties are used to label two or more different molecular probes used in an assay. The ability to

differentiate between and/or quantitate each of the independently detectable moieties provides the means to multiplex a hybridization assay because the data that correlates with the hybridization of each of the distinct, independently labeled molecular probe to a particular target sequence can be correlated with the presence, absence or amount of each organism sought to be detected in the sample. Consequently, the multiplex assays of this invention may be used to simultaneously or sequentially detect the presence, absence or quantity of two or more organisms in the same sample and in the same assay.

Unlabeled Molecular Probes:

The molecular probes that are used for the practice of this invention need not be labeled with a detectable moiety to be operable within the methods of this invention. It is possible to detect the probe/target sequence complex formed by hybridization of the probing nucleobase sequence of the probe to the target sequence using an antibody raised to bind to the probe/target sequence complex. As a non-limiting example, a PNA/nucleic acid complex may be detected using an antibody that specifically interacts with the complex, under suitable antibody binding conditions. Suitable antibodies to PNA/nucleic acid complexes and methods for their preparation and use are described in WIPO Patent Application WO95/17430 as well as US 5,612,458, herein incorporated by reference. Similarly, antibodies to DNA/DNA hybrids are well known in the art and can be made and used as described in US 5,200,313, herein incorporated by reference.

Self-Indicating "Beacon" Probes:

The labels attached to "Beacon" probes comprise a set (hereinafter "Beacon Set(s)") of energy transfer moieties having at least one energy transfer donor and at least one energy transfer acceptor moiety. Typically, the Beacon Set will include a single donor moiety and a single acceptor moiety. Nevertheless, a Beacon Set may contain more than one donor moiety and/or more than one acceptor moiety. The donor and acceptor moieties operate such that one or more acceptor moieties accepts energy transferred from the one or more donor moieties or otherwise quenches the signal from the donor moiety or moieties. Though the previously listed fluorophores (with suitable spectral properties) might also operate as energy transfer acceptors, preferably, the acceptor moiety is a quencher moiety. Preferably, the quencher moiety is a non-fluorescent

aromatic or heteroaromatic moiety. The preferred quencher moiety is 4-((4-(dimethylamino)phenyl)azo) benzoic acid (dabcyl).

Transfer of energy between donor and acceptor moieties of a "Beacon" probe may occur through collision of the closely associated moieties of a Beacon Set(s) or through a non radiative process such as fluorescence resonance energy transfer (FRET). For FRET to occur, transfer of energy between donor and acceptor moieties of a Beacon Set requires that the moieties be close in space and that the emission spectrum of a donor(s) have substantial overlap with the absorption spectrum of the acceptor(s) (See: Yaron et al. *Analytical Biochemistry*, 95: 228-235 (1979) and particularly page 232, col. 1 through page 234, col. 1). Alternatively, collision mediated (radiationless) energy transfer may occur between very closely associated donor and acceptor moieties whether or not the emission spectrum of a donor moiety(ies) has a substantial overlap with the absorption spectrum of the acceptor moiety(ies) (See: Yaron et al., *Analytical Biochemistry*, 95: 228-235 (1979) and particularly page 229, col. 1 through page 232, col. 1). This process is referred to as intramolecular collision since it is believed that quenching is caused by the direct contact of the donor and acceptor moieties (See: Yaron et al.).

(i) *Linear Beacons:*

In a preferred embodiment, the self-indicating "Beacon" probe is a Linear Beacon as more fully described in co-pending and commonly owned patent application USSN 09/179,162 (now allowed), entitled: "Methods, Kits And Compositions Pertaining To Linear Beacons", herein incorporated by reference.

(ii) *PNA Molecular Beacons:*

In a preferred embodiment, the self-indicating "Beacon" probe is a PNA Molecular Beacon as more fully described in co-pending patent application: USSN 09/179,298, entitled: "Methods, Kits And Compositions Pertaining To PNA Molecular Beacons", herein incorporated by reference.

(iii) *DNA Molecular Beacons:*

In a preferred embodiment, the self-indicating "Beacon" probe is a nucleic acid molecular beacon as more fully described in US 5,925,517, entitled: "Detectably Labeled Dual Conformation Oligonucleotide Probes, Assays and Kits".

Detecting Energy Transfer:

Hybrid formation of a self-indicating "Beacon" probe with a target sequence can be monitored by measuring at least one physical property of at least one member of the Beacon Set that is detectably different when the hybridization complex is formed as compared with when the "Beacon" probe exists in the absence of target sequence. We refer to this phenomenon as the self-indicating property of "Beacon" probes. This change in detectable signal results from the change in efficiency of energy transfer between the donor and acceptor caused by hybridization of the "Beacon" probe to the target sequence. Preferably, the means of detection will involve measuring fluorescence of a donor or acceptor fluorophore of a Beacon Set. Most preferably, the Beacon Set will comprise at least one donor fluorophore and at least one acceptor quencher such that the fluorescence of the donor fluorophore is used to detect, identify or quantitate hybridization of the probe to the target sequence.

Other Self-Indicating Probes:

In another embodiment, the self-indicating probes of this invention are of the type described in WIPO patent application WO97/45539. The self-indicating probes described in WO97/45539 differ as compared with "Beacon" probes in that no quencher or acceptor is required because the reporter group must interact with the nucleic acid to thereby produce a detectable signal. Preferably, the probes of WO97/45539, as used in this invention, are appropriately labeled peptide nucleic acids that produce detectable signal upon hybridization to the target sequence.

Spacer/Linker moieties:

Generally, spacers are used to minimize the adverse effects that bulky labeling reagents might have on hybridization properties of probes. Linkers typically induce flexibility and randomness into the probe or otherwise link two or more nucleobase sequences of a molecular probe. Preferred spacer/linker moieties for the nucleobase polymers of this invention consist of one or more aminoalkyl carboxylic acids (e.g. aminocaproic acid) the side chain of an amino acid (e.g. the side chain of lysine or ornithine) natural amino acids (e.g. glycine), aminooxyalkylacids (e.g. 8-amino-3,6-dioxaoctanoic acid), alkyl diacids (e.g. succinic acid), alkyloxy diacids (e.g. diglycolic acid) or alkyldiamines (e.g. 1,8-diamino-3,6-dioxaoctane). Spacer/linker moieties may also incidentally or intentionally be constructed to improve the water solubility of the molecular probe (For example see: Gildea et al., *Tett. Lett.* 39: 7255-7258 (1998)).

Preferably, a spacer/linker moiety comprises one or more linked compounds having the formula: $-Y-(O_m-(CW_2)_n)_o-Z-$. The group Y is selected from the group consisting of: a single bond, $-(CW_2)_p-$, $-C(O)(CW_2)_p-$, $-C(S)(CW_2)_p-$ and $-S(O_2)(CW_2)_p-$. The group Z has the formula NH, NR^2 , S or O. Each W is independently H, R^2 , $-OR^2$, F, Cl, Br or I; wherein, each R^2 is independently selected from the group consisting of: $-CX_3$, $-CX_2CX_3$, $-CX_2CX_2CX_3$, $-CX_2CX(CX_3)_2$, and $-C(CX_3)_3$. Each X is independently H, F, Cl, Br or I. Each m is independently 0 or 1. Each n, o and p are independently integers from 0 to 10.

Hybridization Conditions/Stringency:

Those of ordinary skill in the art of nucleic acid hybridization will recognize that factors commonly used to impose or control stringency of hybridization include formamide concentration (or other chemical denaturant reagent), salt concentration (i.e., ionic strength), hybridization temperature, detergent concentration, pH and the presence or absence of chaotropes. Optimal stringency for a molecular probe/target sequence combination is often found by the well-known technique of fixing several of the aforementioned stringency factors and then determining the effect of varying a single stringency factor. The same stringency factors can be modulated to thereby control the stringency of hybridization of a PNA to a nucleic acid, except that the hybridization of a PNA is fairly independent of ionic strength. Optimal stringency for an assay may be experimentally determined by examination of each stringency factor until the desired degree of discrimination is achieved.

Suitable Hybridization Conditions:

Generally, the more closely related the background causing nucleic acid contaminants are to the target sequence, the more carefully stringency must be controlled. Blocking probes may also be used as a means to improve discrimination beyond the limits possible by mere optimization of stringency factors. Suitable hybridization conditions will thus comprise conditions under which the desired degree of discrimination is achieved such that an assay generates an accurate (within the tolerance desired for the assay) and reproducible result. Aided by no more than routine experimentation and the disclosure provided herein, those of skill in the art will easily be able to determine suitable hybridization conditions for performing assays utilizing the methods and compositions described herein. Suitable *in-situ* hybridization conditions comprise conditions suitable for performing an *in-situ* hybridization procedure. Thus,

suitable *in-situ* hybridization conditions will become apparent to those of skill in the art using the disclosure provided herein; with or without additional routine experimentation.

Suitable Antibody Binding Conditions:

Suitable antibody binding conditions comprise conditions suitable for binding an antibody to its antigen. Thus, suitable antibody binding conditions will become apparent to those of skill in the art using the disclosure provided herein; with or without additional routine experimentation. By way of general guidance to the practitioner in determining suitable antibody binding conditions, methods for preparing and using antibodies can be found in numerous references including: Molecular Probes Of The Nervous System, Volume 1, "Selected Methods For Antibody and Nucleic Acid Probes", Cold Spring Harbor Laboratory Press, 1993 by S. Hockfield et al.

Harmonization Of Suitable Hybridization Conditions & Suitable Antibody Binding Conditions:

When employing the methods of this invention or in the production of the compositions of this invention, it maybe important to harmonize the hybridization conditions with the antibody binding conditions because the staining of the organisms is performed simultaneously with, or subsequent to, an antibody binding event. Because optimization of the same variables (pH, salt concentration etc.) is involved, aided by no more than routine experimentation, those of skill in the art will easily be able to harmonize the antibody binding conditions and suitable hybridization conditions for performing an assay. It should however be noted that the use of non-nucleic acid, and preferably PNA probes, is preferred when harmonization of the hybridization and antibody binding conditions is required because PNA probe bind more tightly under conditions of physiological salt, conditions under which antibodies are more likely to operate most efficiently.

Blocking Probes:

Blocking probes are nucleic acid or non-nucleic acid probes that can be used to suppress the binding of the probing nucleobase sequence of the molecular probe to a non-target sequence. Preferred blocking probes are PNA probes (See: Coull et al., US 6,110,676, herein incorporated by reference). Typically, blocking probes are closely related to the probing nucleobase sequence and preferably they comprise one or more

single point mutations as compared with the detectable molecular probe that is complementary, or substantially complementary, to the target sequence sought to be detected in the assay. It is believed that blocking probes operate by hybridization to the non-target sequence to thereby form a more thermodynamically stable complex than is formed by hybridization between the probing nucleobase sequence and the non-target sequence. Formation of the more stable and preferred complex blocks formation of the less stable non-preferred complex between the probing nucleobase sequence and the non-target sequence. Thus, blocking probes can be used with the methods and compositions of this invention to suppress the binding of the molecular probe to a non-target sequence that might be present in the organism to be distinguished or otherwise interfere with the performance of the assay. Hence, blocking probes can be used to improve the fidelity of the methods of this invention beyond the limits available using the combined selectivity of binding partners and molecular probes. Blocking probes are particularly advantageous in single point mutation analysis (e.g. single nucleotide polymorphism (SNP) analysis).

Probing Nucleobase Sequence:

The probing nucleobase sequence of a molecular probe is the specific sequence recognition portion of the construct. Therefore, the probing nucleobase sequence is a nucleobase sequence designed to hybridize to a specific target sequence wherein the presence, absence or amount of the target sequence can be used to directly or indirectly detect the presence, absence or number of organisms of interest in a sample.

Consequently, with due consideration to the requirements of a molecular probe for the assay format chosen, the length and sequence composition of the probing nucleobase sequence of the probe will generally be chosen such that a stable complex is formed with the target sequence under suitable hybridization conditions.

Probe Complexes:

In still another embodiment, two probes are designed to, in the aggregate, produce a probing nucleobase sequence that hybridizes to the target sequence sought to be detected and thereby generates a detectable signal whereby the nucleobase sequence of each individual molecular probe comprises half or approximately half of the complete complement to the target sequence. As a non-limiting example, the nucleobase sequences of the two probes might be designed using the assay as described in European

Patent Application 849,363, entitled: "Method of identifying a nucleic acid using triple helix formation of adjacently annealed probes" by H. Orum et al. (See: EPA 849,363). Similar compositions comprising a PNA probe triplex have been described in copending and commonly owned application USSN 09/302,238, herein incorporated by reference (also published as WO99/55916). Using this methodology, the probes that hybridize to the target sequence may or may not be labeled since it is the probe complex formed by the annealing of the adjacent probes that is directly detected and not the probes that directly bind to the target sequence.

Organism Of Interest:

The assay user or designer will select the organism of interest. As used herein, the organism of interest is a microorganism, tissue or microscopic sized cell. The organism of interest may be a cell, bacteria, virus, yeast, fungi, other unicellular organism or a multicellular organism. Thus, there are no limitations in the organism of interest except that it be microscopic in size.

The organism of interest is generally selected to be an organism characterized by domain, kingdom, group, class, genus, species, taxon, subclass, subspecies, serotype, strain or by any other recognized means of characterization of the organism of interest. Optionally, but not necessarily, the organism of interest will be chosen such that it is to be distinguished from a closely related organism or organisms wherein an antibody or probe based assay, alone, is not adequate to properly characterize the organism of interest from the organism or organisms to be distinguished.

Determining Particles Or Beads:

In certain embodiments of this invention, particles or beads are used as a solid carrier in the assay methods wherein organisms bound to the particles or beads are determined. In certain of these embodiments, it is further required that a property inherent to the particle composition be independently determined. For these assays, coded beaded supports are used.

Coded Beaded Supports:

Coded beaded supports are particles or beads that possess an inherent independently detectable property that can be determined independently of the properties of constituents that are bound to the particle. Hence, many different types of coded beaded supports can be made independently detectable so that an assay can be

designed to assign a parameter of interest to each different particle or bead type and thereby provide a way to determine that parameter by determining the particle type. Non-limiting examples of parameters that may be associated with (i.e. used to "code") the particle type include the sample source or another independent assay parameter.

5 It is not intended that the coded beaded supports must be determined by instrument where the coded beaded supports can be determined without the aid of an instrument. For example, four coded beaded supports might comprise a visible red, blue, green and yellow color. Because these coded supports are visibly colored, they can be determined by visual analysis, or if desired, by instrument analysis.

10 Particle Sorters And Sorting Particles:

In certain embodiments of this invention, it is required that particles or beads be sorted and that constituents thereon be determined. Any flow cytometer is a particle sorter suitable for determining characteristics of the constituents of micro particles or micro beads provided that the assay need not determine a coded beaded support. If the
15 assay must determine a coded beaded support then the flow cytometer must be so equipped to make that determination unless the determination can be made visually or in another way.

As discussed in the Background section above, the Luminex 100™ represents an instrument capable of both determining the coded beaded support as well as
20 determining properties of constituents bound to said particle. Thus, the instrument provides a means to multiplex assays such that each different type of coded beaded support can be used to code for a parameter (property) of interest or sample source and wherein the constituent or constituents bound to the particles or beads are analyzed simultaneously or sequentially for an independent property or properties. In the context
25 of the present invention, the constituents bound to the particles are organisms or cells. It is noted that Luminex does not appear to teach or suggest that their coded beaded supports and/or instruments are suitable for the direct analysis of organisms or cells immobilized to said coded beaded supports.

Microscope Slides:

30 In other preferred embodiments of this invention, the solid carrier is chosen to be a microscope slide. The microscope slide may be made of glass, quartz, plastic or other material that is translucent to light. Microscope slides are a useful solid carrier because

the surface can be easily modified with a binding partner and then stained organisms, which are immobilized to the solid carrier, can be viewed in an appropriately equipped microscope. In certain preferred embodiments, a microscope slide can be manufactured to be an array by the application of the same or different binding partners at each of the unique, identifiable locations.

Arrays:

In other preferred embodiments of this invention, the solid carrier is chosen to be an array. Arrays comprise unique, identifiable locations that differ in a way that can be used to determine a characteristic or property of interest. In the context of the present invention, the same or a different binding partner will be linked to each of the unique, identifiable locations of the array. In a preferred embodiment, the different binding partners are chosen to link a different organism of interest to each of the unique, identifiable locations of the array.

Immobilizing Binding Partners To The Solid Carrier:

Numerous surface chemistries exist for the modification of surfaces for the purpose of producing surface bound functional groups to which binding partners can be linked for immobilization. Additionally, numerous solid carriers possessing reactive functional groups are commercially available. These include the coded beaded supports available from Luminex (See Luminex Product Literature), chromatographic packing materials that are available from vendors such as Amersham Pharmacia (See Amersham Pharmacia Catalog) and coated glass slides that are available from Corning and can be used to fashion micro arrays. (See: Corning MicroArray technology at www.corning.ny.com) Once the solid carrier is derivatized to possess available functional groups, the binding partner is easily reacted with the surface bound functional group to thereby effect binding partner immobilization. Those of ordinary skill in the art will further appreciate that any solid carrier can be derivatized using available surface chemistry technology to fashion a custom solid carrier bearing the desired immobilized binding partner or partners. Hence, this invention is not intended to be limited by any particular commercially available solid carrier or any particular surface chemistry modification.

Advantages Of The Present Invention:

It is an advantage of the present invention that the selectivity and discriminating power of binding partners, as used in a capture assay, is combined with the selectivity and discriminating power of molecular probes, as used to stain organisms, tissues or cells, in a way that provides for two independent levels of certainty and/or discrimination in the assay. Hence, the assays and compositions of this invention provide for either or both a greater ability to discriminate organisms, tissues and/or cells and/or a greater deal of certainty with respect to the result of the assay.

II. Preferred Embodiments of the Invention:

a. Assay Methods:

(i) *Method For Determining An Organism*

In one embodiment, this invention is directed to a method for determining an organism of interest in a sample from another organism or organisms to be distinguished. The method comprises treating the sample, or a portion thereof, with at least one detectable molecular probe wherein the molecular probe or probes are selected such that either: (i) both the organism of interest and the other organism or organisms react with the molecular probe in a way that produces detectable organisms of interest and a detectable other organism or organisms to be distinguished; or (ii) only the organism of interest reacts with the molecular probe in a way that produces only detectable organisms of interest. According to the method, the sample, or a portion thereof, is also contacted with a solid carrier to which has been immobilized a binding partner such that if (i) applies then the binding partner is chosen to be reactive only with the detectable organism of interest but not reactive with the detectable other organism or organisms to be distinguished; but if (ii) applies then the binding partner is chosen to be generally reactive with the detectable organism of interest but also may be reactive with the other organism or organisms to be distinguished. Once the organisms have been stained and immobilized to the solid carrier, the method involves determining the presence, absence, position or number of detectable organisms immobilized to the solid carrier and correlating the result with the presence, absence, or number of the organisms of interest in the sample, or portion thereof.

In determining the organism of interest, the method requires that a correlation between the presence, absence, position or number of detectable organisms immobilized

to the solid carrier be made with the presence, absence or number of organisms of interest in the sample, or portion thereof. This correlation is straight forward since the organisms that are determined by operation of the method come directly from the sample, or portion thereof.

5 In preferred embodiments of this method, the molecular probe stains all organisms of a domain, kingdom, group, class, genus, species, taxon, subclass, subspecies, serotype or strain without regard to whether or not this represents the organism of interest. For this preferred embodiment, it is the binding partner that is specific for the domain, kingdom, group, class, genus, species, taxon, subclass,
10 subspecies, serotype or strain that is the organism of interest.

As a non-limiting example of this embodiment, the molecular probe is selected to be suitable for staining all bacteria in the sample wherein the organism of interest is Salmonella bacteria. Thus, the use of a solid carrier immobilized binding partner that is an antibody to Salmonella bacteria will facilitate, for determination, the selective
15 immobilization of only the stained Salmonella bacteria. Since the other organisms are not immobilized to the solid carrier, they are not determined by the method even if they are detectably stained. Hence, it is the interaction with the solid carrier that provides the ultimate selectivity for the organism of interest in this embodiment.

In another preferred embodiment of this method, the molecular probe stains only the domain, kingdom, group, class, genus, species, taxon, subclass, subspecies, serotype
20 or strain that is the organism of interest. For this preferred embodiment, the binding partner is chosen to bind a particular domain, kingdom, group, class, genus, species, taxon, subclass, subspecies, serotype or strain without regard to whether or not this represents the organism of interest, provided however that the organism of interest is
25 within the scope of the binding capability of the binding partner.

As a non-limiting example of this embodiment, the molecular probe is selected to be specific for the staining of Salmonella bacteria wherein Salmonella bacteria represents the organism of interest. Thus, the use of a solid carrier immobilized binding partner that is an antibody to all bacteria will facilitate, for determination, the selective
30 immobilization of all bacteria. Although other bacteria may be captured by the solid carrier, they are not detectably labeled and are thus, not determined by the method.

Hence, it is the presence of the stain that provides the ultimate selectivity for the organism of interest in this embodiment.

In yet another preferred embodiment of this method, the molecular probe stains only the domain, kingdom, group, class, genus, species, taxon, subclass, subspecies, serotype or strain that is the organism of interest. For this preferred embodiment, the binding partner is also specific for only the domain, kingdom, group, class, genus, species, taxon, subclass, subspecies, serotype or strain that is the organism of interest.

As a non-limiting example of this embodiment, the molecular probe could be selected to be specific for the staining of Salmonella bacteria wherein Salmonella bacteria represents the organism of interest. Additionally, the use of a solid carrier immobilized binding partner that is an antibody that is specific to Salmonella bacteria will facilitate, for determination, the selective immobilization of only Salmonella bacteria. In this way, the specificity is achieved both at the level of staining using the molecular probe and again at the level of capture using the antibody. Hence, in this embodiment, the assay provides for certainty of the result at two different levels of molecular discrimination.

It should be noted that certainty at two levels of molecular discrimination is very useful since although binding partners and molecular probes are designed to be selective, they cannot be tested for cross reaction against all organisms or other interfering matter. Hence, even if some level of cross reactivity occurs in the assay with either of the molecular probe or binding partner, because the selectivity substantially differs at each different level of molecular discrimination, it is not likely that a particular cross reacting species will exhibit cross reactivity at both levels of discrimination. Consequently, the certainty of a result is significantly increased where a positive result requires, as does certain embodiments of this invention, that the assay perform an interrogation at two levels of molecular discrimination.

(ii) *Method For Sorting And Determining An Organism Using Coded Beaded Supports*

In another embodiment, this invention is directed to a method for sorting and determining an organism or organisms of interest in a sample or samples using coded beaded supports. The method comprises treating the sample, or a portion thereof, with one or more detectable or independently detectable molecular probes wherein the one or more molecular probes are selected such that either: (i) the detectable probe or probes react with the different organisms to be determined in a way that produces different detectable organisms that possess the same stain; or (ii) the independently detectable probes react with the different organisms to be determined in a way that produces different independently detectable organisms that possess an independently detectable stain. According to the method, the sample, or a portion thereof, is also contacted with one or more different types of coded beaded supports, wherein each different type of coded beaded support can be independently determined in a suitable particle sorter and wherein to each different type of coded beaded support has been immobilized a particular binding partner that is chosen to select a particular organism or organisms such that detectable or independently detectable organisms become selectively bound to the coded beaded supports as a result of the occurrence of specific binding partner interactions. According to the method, the different types of coded beaded supports are then sorted in a suitable particle sorter. The presence, absence, or number of the detectable organism or organisms, or each of the independently detectable organisms, immobilized to each different type of coded beaded support is also determined. This result is then either: (iii) correlated with the code that is associated with a particular immobilized binding partner to thereby determine the presence, absence or number of each of the different organisms of interest in the sample or portion thereof; or (iv) correlated with the code for a sample source from which the sample, or portion thereof, was derived to thereby determine the presence, absence or number of each of the different organisms of interest in each different sample, or portion thereof.

In determining the organism or organisms of interest, the present method requires that a correlation between the presence, absence, or number of detectable organisms immobilized to the coded beaded support be made with the presence, absence or number of organisms of interest in the sample, or portion thereof. This

correlation is straight forward since the organisms that are determined by operation of the method come directly from the sample, or portion thereof.

However, certain embodiments of the present method also require that a correlation be made between the "code" of the coded beaded support and a particular binding partner. Since the coded beaded supports are manufactured, the identity and properties of the binding partner immobilized thereto is predetermined and known. Consequently, this correlation is again straight forward.

Additionally, certain other embodiments of the present method require that a correlation be made between the "code" of the beaded support and the source of a particular sample, or portion thereof. Because for this embodiment each sample is assigned to a particular coded support and because the "code" for each sample is predetermined and known, the resulting correlation is also straight forward.

In one preferred embodiment of this method, the detectable molecular probe or probes stain the organism or organisms of interest with the same stain. For this embodiment, the binding partner that is associated with each different type of coded beaded support is chosen to be specific for one of the different organisms of interest such that the sorting of the different types of coded beaded supports determines each of the different organisms of interest in a sample, or portion thereof, based upon the identity of each different binding partner that is associated with each particular coded beaded support.

As a non-limiting example of this embodiment, the molecular probe is chosen to stain all bacteria in the sample, or portion thereof, wherein the organism of interest for one of the coded beaded supports is Salmonella. Thus, the use of a coded beaded support (Bead Type 1) to which is linked an antibody that is specific to Salmonella bacteria, will facilitate the selective immobilization of only the stained Salmonella bacteria to Bead Type 1. Since this coded beaded support is associated only with the presence of the Salmonella antibody, a determination of the code of the bead (Bead Type 1), in combination with a determination of stained organisms on said bead, provides all the information needed to make a determination of presence, absence or number of Salmonella bacteria in the sample.

It should be noted that the same sample can also be analyzed for other bacteria such as, for example, *Pseudomonas aeruginosa*. According to the method, a second

coded beaded support (Bead Type 2), to which is linked an antibody that is specific to *Pseudomonas aeruginosa* bacteria, will facilitate the selective immobilization of only the stained *Pseudomonas aeruginosa* bacteria to Bead Type 2. Since all the bacteria of the sample, or portion thereof, are stained with the same color, a determination of the "code" of the bead (Bead Type 2) in combination with a determination of stained organisms on the bead, provides all the information needed to make a determination of the presence, absence or number of *Pseudomonas aeruginosa* bacteria in the sample. Hence, the method provides for multiplex analysis since many different coded beaded supports can be used for the analysis of numerous different organisms of interest that might be present in the same sample.

Moreover, it will also become apparent to the ordinary practitioner that the number of different organisms of interest that can be detected from the same sample using this methodology is limited only by the availability of different bead types and the availability of specific binding partner complexes. Nevertheless, even this is not a true limitation since antibodies can be raised to most antigens and coded beads can be custom made.

In another preferred embodiment of this method, the independently detectable molecular probe or probes stain all of the organisms of interest provided that some or all of the different organisms of interest are stained differently. For this embodiment, each binding partner associated with each different type of coded beaded support is chosen to select among the same or differently stained organisms such that the sorting of the different types of coded beaded supports, when considered in combination with the stain of the organism or organisms bound to each different type of coded beaded support, is used to determine each of the different organisms of interest in a sample, or portion thereof.

As a non-limiting example of this embodiment, each of the independently detectable molecular probes are selected to be specific for staining different organisms of interest such that the different organisms of interest are independently detectable when stained. For example, the different organisms of interest are *Salmonella* bacteria, *E. coli* bacteria and *Pseudomonas aeruginosa* bacteria wherein the bacteria are stained, using suitable fluorophore labeled molecular probes, with red, green and blue fluorophores, respectively. For this example, the binding partner is selected to be: (a) suitable for

binding the bacteria to the same coded beaded support; or (b) suitable for binding individual bacterial strains to the same or different coded beaded supports. As evident by the example described below, the assay can be utilized whereby both (a) and (b) apply. Thus, it is not clearly not a limitation of this embodiment of the invention that
 5 either (a) or (b) apply.

For simplicity, assume that the binding partner on Bead 1 is specific for *E. coli* and the binding partner on Bead 2 is specific for both *Salmonella* bacteria and *Pseudomonas aeruginosa* bacteria. In this way both conditions (a) and (b) are generally met. Since the color of the organisms bound to the solid carrier provides one level of selectivity, and the
 10 nature of the binding partner associated with different coded beaded supports provides a second level of selectivity, a determination of both the color or colors of organisms immobilized to the beads and the "code" of each bead upon which the color determination has been made, provides all the information necessary to determine the presence, absence or amount of the different organisms in the sample, or portion
 15 thereof.

More specifically, if situation (a) applies the presence, absence or quantity of red, green or blue fluorophore detected on the only type of coded beaded support determines the presence, absence or number of *Salmonella* bacteria, *E. coli* bacteria and *Pseudomonas aeruginosa* bacteria in the sample, or portion thereof. Consequently, for
 20 this embodiment, the ultimate selectivity is achieved using the molecular probe. However, if situation (b) applies, then the beads can be interrogated for extraneous colors. In the present example, Bead 1 need only be analyzed for green (*E. coli*), and Bead 2 need only be analyzed for red (*Salmonella* bacteria) and blue (*Pseudomonas aeruginosa* bacteria). Hence, in this embodiment of a multiplex assay, selectivity is
 25 achieved at both the level of interaction of the molecular probe and again at the level of the binding partner as determined by the nature of each different coded beaded support. Therefore, the certainty of the result is improved in situation (b).

In yet another preferred embodiment of this method, the independently detectable molecular probes stain all of the organisms of interest differently. For this
 30 embodiment, the binding partner associated with each different type of coded beaded support is the same. However, each different coded beaded support "codes" for a different sample such that the determination of the stain or stains on each different coded

beaded support specifically determines each of the different organisms of interest in the sample, or portion thereof, and each different coded beaded support identifies the source of the sample, or portion thereof. Hence, the determination of the color of organisms on each coded beaded support, when considered with the bead "code", provides all the information needed to determine the organisms in the sample and the sample source, respectively.

As a non-limiting example of this embodiment, 100 different coded beaded supports are all derivatized with an antibody or antibodies that is/are suitable for the capture of 5 different organisms of interest. To each sample to be tested is added five different fluorophore labeled molecular probes under conditions that will, if present, stain each of the five different bacteria of interest one of either, blue, yellow, orange, red or green. To each of 100 different samples that are to be tested is added one of the 100 different coded beaded supports such that all 100 samples are identifiable by a different one of the 100 different coded beaded supports. Since each coded beaded support now "codes" for a different sample, the coded beaded supports can be analyzed simultaneously (in a mixture) or sequentially to thereby obtain the result for the many different samples, or portions thereof. Consequently, this embodiment of the method pertains to multiplexing samples and optionally multiplexing the analysis of sample analytes; it being noted that the sample analytes (each of the five different organisms of interest) are multiplexed by using the five different independently detectable molecular probes and each different sample source being multiplexed using independently determinable coded beaded supports. Hence, in this example, multiple levels of multiplexing of the assay is performed such that throughput of the system is substantially improved.

It should be noted, that the number of different organisms of interest that can be determined is limited only by the number of different independently detectable molecular probes that can be prepared and that five organisms per sample is not intended to be a limit on the method. Furthermore, it is evident that 100 different types of coded beaded supports is not intended to be a limit on the method.

(iii) *Method For Sorting And Determining An Organism Using An Array*

In yet another embodiment, this invention is directed to a method for sorting and determining different organisms of interest in a sample using an array. The method comprises treating the sample, or a portion thereof, with one or more detectable or independently detectable molecular probes wherein the one or more molecular probes are selected such that either: (i) the detectable probe or probes react with the different organisms to be determined in a way that produces different detectable organisms that possess the same stain; or (ii) the independently detectable probes react with the different organisms to be determined in a way that produces different independently detectable organisms that possess an independently detectable stain. According to the method, the sample, or a portion thereof, is contacted with a solid carrier array to which binding partners have been immobilized at unique, identifiable locations such that the detectable or independently detectable organisms are selectively bound to the locations on the array as a result of the occurrence of specific binding partner interactions. The presence, absence or number of the detectable or independently detectable organisms immobilized at the many different locations of the array is then determined and the result is correlated with the particular binding partner immobilized to each location on the array to thereby determine the presence, absence or number of the different organisms of interest in the sample.

In determining the organism of interest, the method requires that a correlation between the presence, absence, position or number of detectable organisms immobilized to the solid carrier be made with the presence, absence or number of organisms of interest in the sample, or portion thereof. This correlation is straight forward since, although the array provides for sorting, the organisms that are determined by operation of the method come directly from the sample, or portion thereof.

In one embodiment of this method, the detectable molecular probe or probes stain all of the different organisms with the same stain. For this embodiment, the binding partner is chosen to be specific for each of the different organisms of interest such that the sorting of the organisms on the array resulting from the binding partner interactions occurring at the unique locations is used to thereby determine each of the different organisms of interest in the sample, or portion thereof, based solely upon the identity of the different binding partners at the unique locations. Because the array is

fabricated, the identity of each binding partner at a unique, identifiable location is predetermined and known.

As a non-limiting example of this embodiment, a molecular probe is chosen to stain all bacteria in the sample with a red fluorophore. Additionally, an array of three different binding partners is prepared such that at each unique, identifiable location (a "spot") on the array, a different one of three binding partners, to each of *Salmonella* bacteria, *E. coli* bacteria and *Pseudomonas aeruginosa* bacteria, is immobilized. Thus, the array can be said to comprise a *Salmonella* spot, an *E. coli* spot and a *Pseudomonas aeruginosa* spot. Consequently, a determination of the red fluorophore at one or more of the "spots" on the array, when correlated with the identity of the binding partner immobilized at the unique identifiable location, provides the information necessary to determine the presence, absence or amount of each of *Salmonella* bacteria, *E. coli* bacteria and *Pseudomonas aeruginosa* bacteria in the sample, or portion thereof.

In still another embodiment of this method, the independently detectable molecular probe or probes stain all of the organisms of interest provided that some or all of the different organisms of interest are stained differently. For this embodiment, each binding partner associated with a unique location on the array is chosen to select among the same or differently stained organisms such that the sorting of the organisms on the array resulting from the binding partner interactions occurring at the unique locations, when considered in combination with the stain of the organism or organisms bound to each unique location, is used to determine each of the different organisms of interest in the sample, or portion thereof.

As a non-limiting example of this embodiment, two or more independently detectable molecular probes are chosen to stain *Salmonella* bacteria, *E. coli* bacteria and *Pseudomonas aeruginosa* bacteria in the sample, or portion thereof with a red, green or blue fluorophore, respectively. Additionally, an array of two different binding partners is prepared such that at each of two unique, identifiable locations (a "spots") on the array, *Salmonella* bacteria and *E. coli* bacteria bind to the binding partner at spot 1 and *Pseudomonas aeruginosa* bacteria binds to the binding partner at spot 2. Thus, a determination of a red, green or blue fluorophore at each "spot" on the array, when correlated with the identity of the binding partner immobilized at the unique, identifiable location, provides the information necessary to determine the presence, absence or

amount of each of Salmonella bacteria, E. coli bacteria and Pseudomonas aeruginosa bacteria in the sample, or portion thereof. In particular, a red (Salmonella) and/or green (E. coli) signal should be determined at spot 1 and a blue (Pseudomonas aeruginosa) signal should be determined at spot 2. It should be noted that in this embodiment of the method, the certainty of the assay is increased because the selectivity for some of the organisms is achieved at both the level of the molecular probe and at the level of the chosen binding partner. Hence, this is yet another example of the flexibility in multiplexing that is available with the presently described inventive methods.

(iv) *General Attributes Of The Aforementioned Methods:*

According to the methods, the detectable molecular probe is selected from the group consisting of a nucleic acid and a non-nucleic acid oligomer. In preferred embodiments, the non-nucleic acid is a peptide nucleic acid oligomer.

According to the method, the detectable molecular probe need not be labeled with a detectable moiety. For these embodiments, the detectable molecular probe may be detected using a detectable antibody that specifically binds to a detectable molecular probe/target sequence complex. Preferably for these embodiments, the detectable molecular probe is an unlabeled peptide nucleic acid and the peptide nucleic acid/target sequence is detected with a suitable detectable antibody.

In other preferred embodiments of this invention, the detectable molecular probe is labeled with a detectable moiety. The preferred detectable moiety suitable for the practice of this invention include: a chromophore, a fluorochrome, a spin label, a radioisotope, an enzyme, a hapten and a chemiluminescent compound.

It is generally not important to the success of any of the aforementioned methods whether or not the staining with molecular probes is performed before, during or after the immobilization of the particle, cell or tissue to the solid carrier provided however that both steps are performed before the determination of organism or organisms is made. Thus, in one embodiment the sample, or portion thereof, is treated with the detectable molecular probe or probes before being contacted with the solid carrier. In another embodiment, the sample, or portion thereof, is contacted with the solid carrier before being treated with the detectable molecular probe or probes. In yet another

embodiment, the sample, or portion thereof, is simultaneously contacted with both the solid carrier and treated with the detectable molecular probe or probes.

b. Compositions:

5 In still another embodiment, this invention is directed to a composition comprising one or more organisms stained with one or more detectable molecular probes and a solid carrier to which is immobilized an binding partner. The one or more organisms are linked to the solid carrier through the interaction of the organism and its binding partner. For this embodiment, preferably the binding partner is an antibody
10 and the organism is the antigen to the antibody. In other preferred embodiments, the solid carrier is a solid carrier array comprising antibodies to many different organisms of interest which have each been immobilized at unique identifiable locations on the array. Alternatively, the solid carrier is a coded beaded support, a microscope slide or a membrane.

15 In still another embodiment, this invention is directed to a composition comprising two or more different organisms of interest that are detectably or independently stained with one or more molecular probes and a mixture of two or more different types of coded beaded supports. To each different type of coded beaded support has been immobilized a different binding partner that is selected to detect a
20 particular organism of interest and wherein the different detectable or independently detectable organisms are selectively bound to the coded beaded supports as a result of the occurrence of specific binding interactions of the binding partner and the organisms. For this embodiment, preferably the binding partner is an antibody and the organism is the antigen to the antibody.

25 In yet another embodiment, this invention is directed to a composition comprising two or more different organisms of interest that are detectably or independently stained with one or more molecular probes and a solid carrier array to which binding partners have been immobilized at unique identifiable locations such that the detectably or
independently stained organisms are selectively bound to the locations on the array as a
30 result of the occurrence of specific binding partner interactions. For this embodiment, preferably the binding partner is an antibody and the organism is the antigen to the antibody.

Having described the preferred embodiments of the invention, it will now become apparent to one of skill in the art that other embodiments incorporating the concepts described herein may be used. It is felt, therefore, that these embodiments
5 should not be limited to disclosed embodiments and examples but rather should be limited only by the spirit and scope of the following claims.

Examples:

This invention is now illustrated by the following examples that are not intended to be limiting in any way.

5 Example 1:PNA Oligomers As Molecular Probes:

PNA Oligomers were prepared from commercial reagents and instrumentation obtained from Applied Biosystems, Foster City, CA using well known methods.

10 *PNA Oligomers Prepared:*

Table 1		
Probe ID	PNA Probe Sequence	Seq. Id. No.
Bac Uni	Flu-OO-CTG-CCT-CCC-GTA-GGA-NH ₂	1

All PNA sequences are written from the amine (N-) terminus to the carboxyl (C-) terminus. Flu = 5(6)-carboxyfluorescein; O = 8-amino-3,6-dioxaoctanoic acid

Experimental Methods:15 *Fixation of Cells*

Salmonella choleraesuis and *Listeria monocytogenes* cells were fixed by pelleting 10-20 mL of exponentially growing cultures by centrifugation at 10,000 rpm for 5 minutes. The media was removed and cell pellets were resuspended in an equal volume of Buffer A. Cell suspensions were centrifuged at 10,000 rpm for 5 minutes, the supernatant was removed. Cells were resuspended in an equal volume of Buffer B, incubated at room temperature for 1 hour, pelleted by centrifugation at 10,000 rpm and washed with Buffer A. Final cell pellets were resuspended in an equal volume of Buffer C and stored at -20°C for a minimum of 30 minutes.

25 *Buffer Solutions*

- A 130 mM NaCl, 7 mM Na₂HP0₄, 7 mM NaH₂P0₄, pH 7.0
- B 4% paraformaldehyde in Buffer A
- C 50% ethanol in water
- D 25 mM Tris pH 9.0, 0.5% SDS, 100mM NaCl
- 30 E 10 mM Tris pH 9.0, 1 mM EDTA

Hybridization

For hybridization, 100 μ L of fixed cells were pelleted by centrifugation for 5 minutes at 10,000 rpm, resuspended in 100 μ L of Buffer A, pelleted as described above and finally resuspended in 100 μ L of Buffer D. Fluorescein labeled Bac Uni PNA probe was then added to a final concentration of 300 pmole/mL. Reactions were then incubated at 55°C for 30 minutes. Reactions were pelleted as described above and pellets were washed for 10 minutes at 55°C in 500 μ L Buffer E, followed by centrifugation at 10,000 rpm for 5 minutes. The wash step was repeated twice for a total of 3 washes. Final cell pellets were resuspended in 100 μ L of Buffer E.

Capture Onto Antibody Coated Beads

Two types of coded beads were received from Luminex, one coated with Salmonella-specific antibody (OEM Concepts, Toms River, NJ; the "Salmonella beads") and one with Listeria-specific antibody (OEM Concepts, Toms River, NJ; the "Listeria beads"). A 25 μ L aliquot of each type of coded bead was pelleted by centrifugation and resuspended in 25 μ L of Buffer E. The resuspended beads and the hybridization reactions were combined in equal volumes (5 μ L of each) and incubated at room temperature with shaking for 2 hours. Capture reactions included treatment with *S. choleraesuis* for each coded bead type and treatment with *L. monocytogenes* for each bead type. Reactions were analyzed by spreading 2 μ L of the sample on a preheated microscope slide (~60°C). The slide was then dried on a heat block set at approximately 60°C. A 2 μ L aliquot of mounting media (Vector Laboratories) was then added and a coverslip applied. All slides were then examined microscopically using a 60X objective and a double band pass filter (FITC/Texas Red).

Results:

With reference to Figure 1, the Salmonella beads (red spheres) effectively captured the green stained *S. choleraesuis* cells (Figure 1A) and did not bind to the green stained *L. monocytogenes* cells (Figure 1B). By microscopic examination, it appears that approximately 25% of the beads were bound to *S. choleraesuis* cells, compared to <1% of beads bound to *L. monocytogenes*. With reference to Figure 2, while the Listeria beads (red spheres) did capture the green stained *L. monocytogenes* cells (Figure 2A), the *S.*

- choleraesuis* cells are also captured by the *Listeria* specific beads (Figure 2B). It is possible that the non-specific binding seen with the *Listeria* beads occurs because the capture conditions were not optimized. Alternatively, the lack of specificity with the *Listeria* beads could be due a lack of specificity of the antibody used to coat the beads. This
- 5 result therefore reinforces why selectivity at multiple levels of molecular discrimination is a preferred means of analysis. Future experimentation is planed to determine why the selected specificity was not achieved with the *Listeria* beads.